

Digital genetics with *aevol*: a view on the (evolutionary) origin of biological complexity

TAGp, Annecy - October 20, 2010

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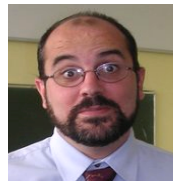
centre de recherche
GRENOBLE - RHÔNE-ALPES

Guillaume Beslon
COMBINING



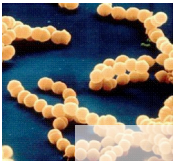
Molecular structure of biological systems

[Koonin, 2009]



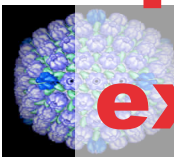
Homo sapiens

~3 billions bp
~25 000 genes



Neisseria meningitidis

~2 millions bp
~2 000 genes

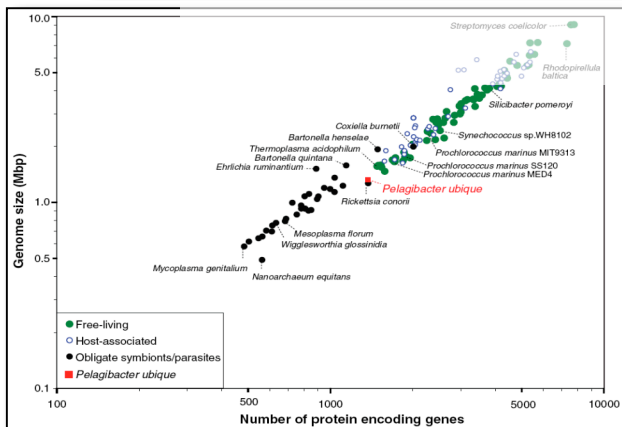
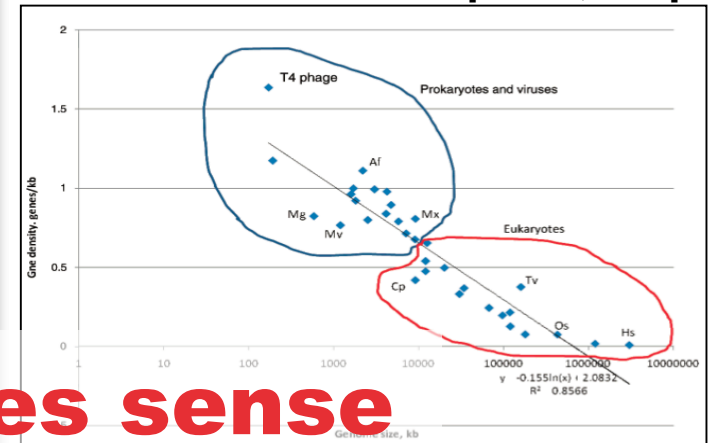


Herpes HSV-1

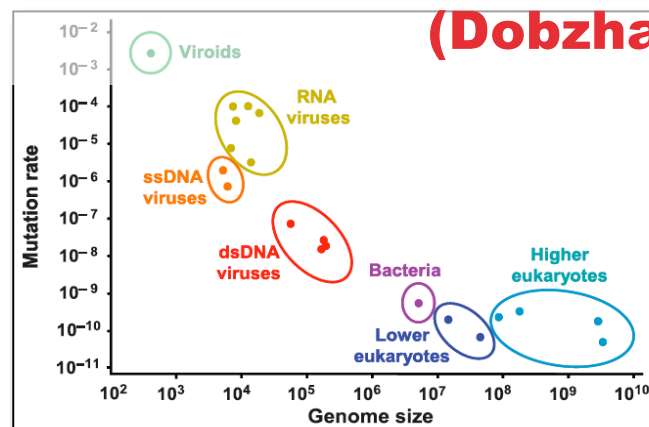
~150 000 bp
~100 genes



“Nothing in biology makes sense except in the light of evolution”

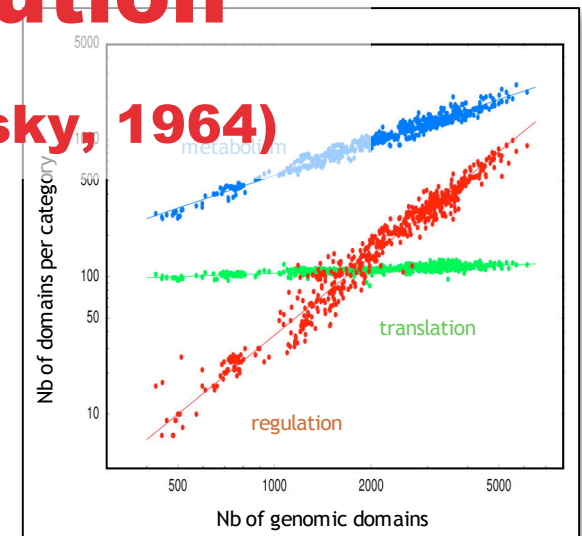


[Giovannoni et al., 2005]



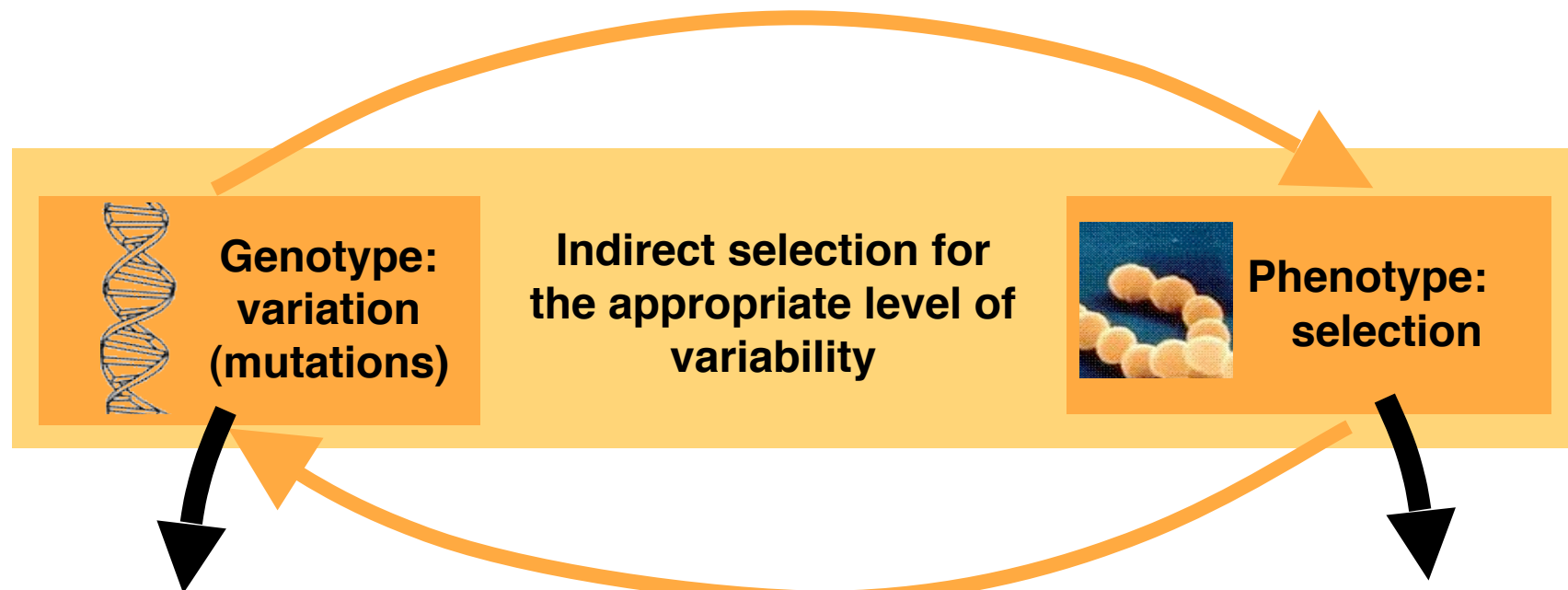
(Dobzhansky, 1964)

[Gago et al., 2010]



[Molina & Van Nimwegen, 2008]

Evolutionary origin of biological systems



Mutational biases:

“Homo Sapiens genome spontaneously undergoes more insertions than deletions”

“Gene duplication spontaneously creates scale-free regulation networks”

Selective costs/advantages:

“In virus and bacteria genomes are physically constrained”

“Complex environments lead to complex regulation networks”



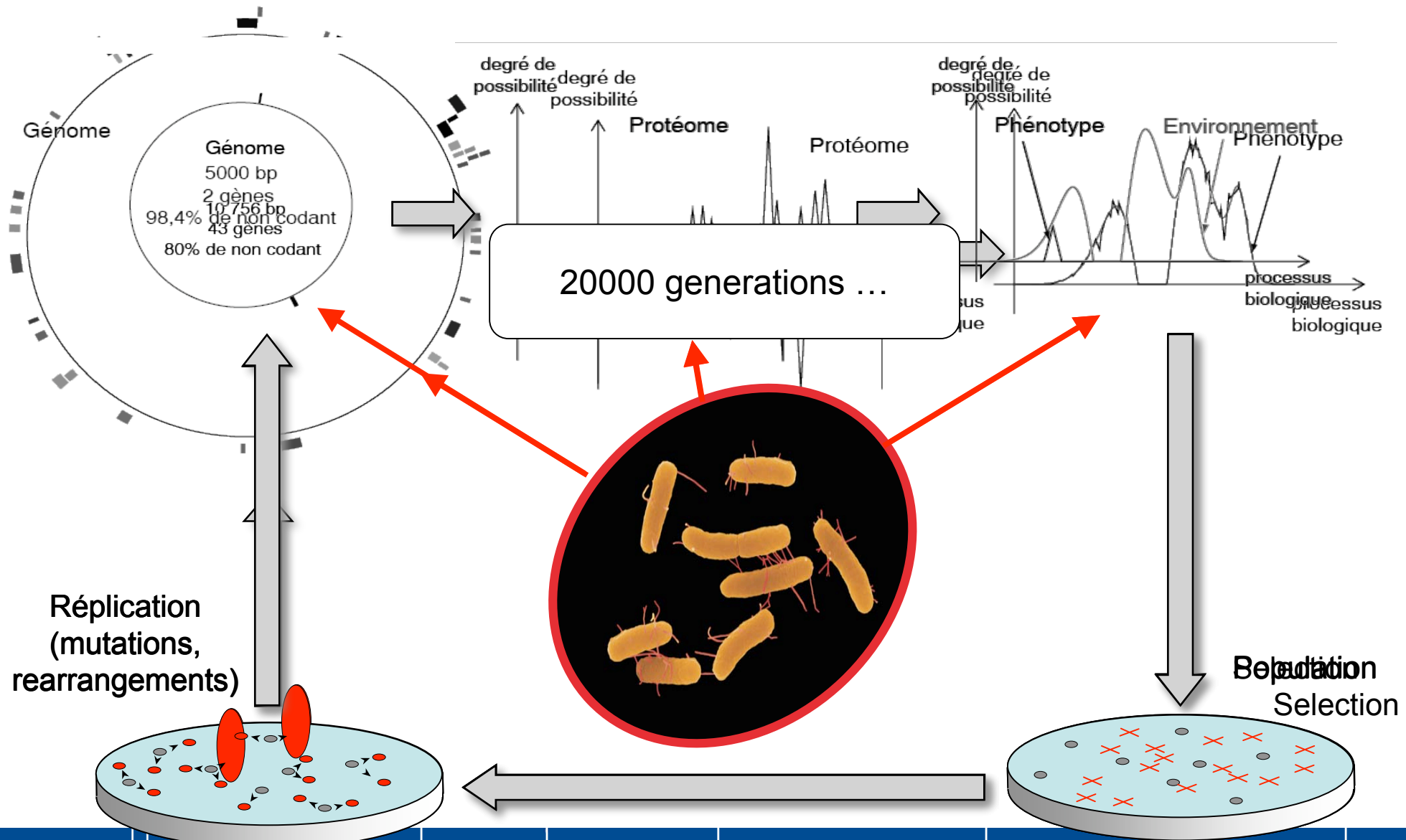
Biological question

1. The scaling of most aspects of genomic architecture and gene structure with genome size fall on a continuum from viruses to prokaryotes to single-celled eukaryotes to multicellular eukaryotes. This suggests that general population-genetic mechanisms, transcending cellular and metabolic features, are the predominant drivers of interspecific divergence in genome architecture. (Lynch, Annu. Rev. Microbiol., 2006)

- Can indirect selection of an appropriate variability level explain the various genetic organizations observed in nature?
 - Mutational burden hypothesis
 - Is there a burden on non-coding region?
- Computational approach
 - Investigations using *in silico* experimental evolution
 - Digital genetics (“real evolution of false organisms”)
 - Individual-Based Modeling of (prokaryotes) evolution



The aevo model (C. Knibbe, 2006)

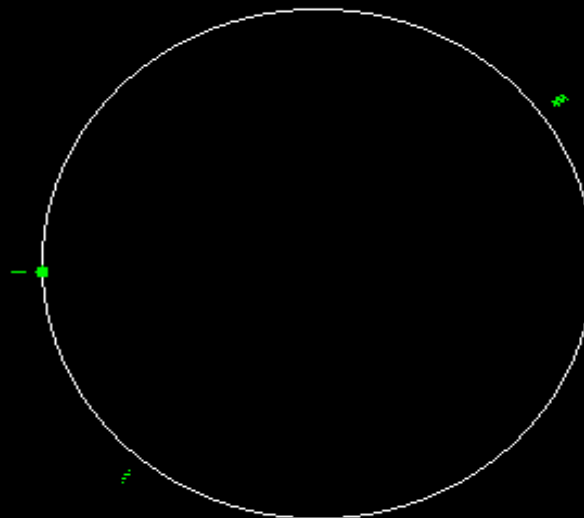


aevol: The movie (« winning » lineage)

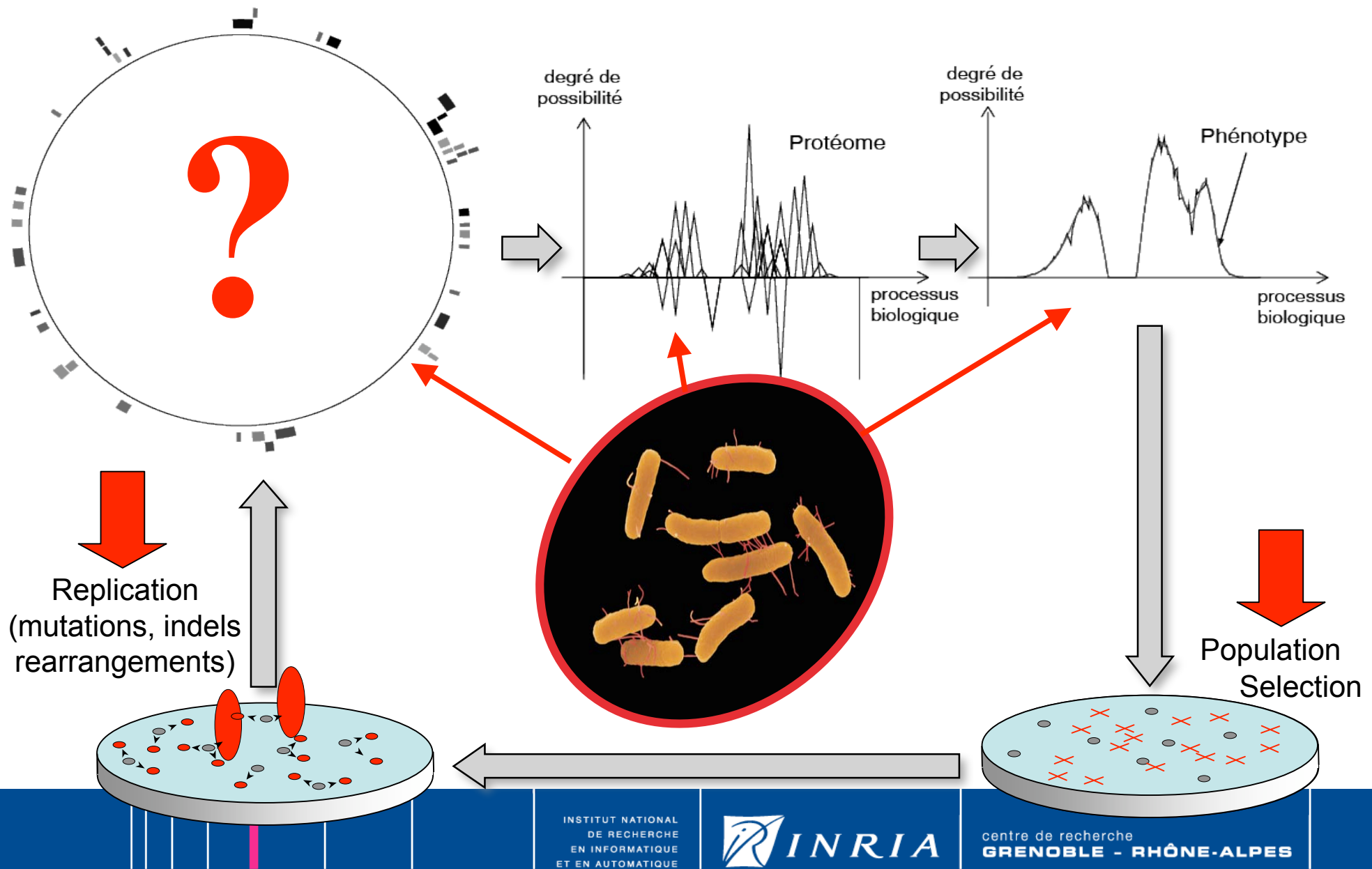
Genome length = 25993 bp

Generation = 129

Small insertion at 19406 of sequence 01



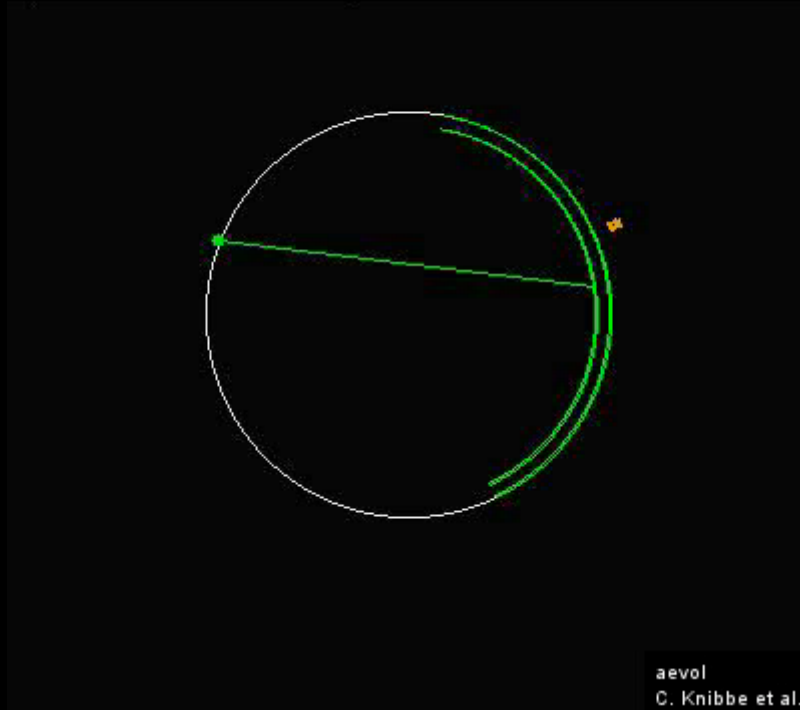
Experimental evolution with aevol



aevol: The movie (II) ...

Genome length = 5000 bp

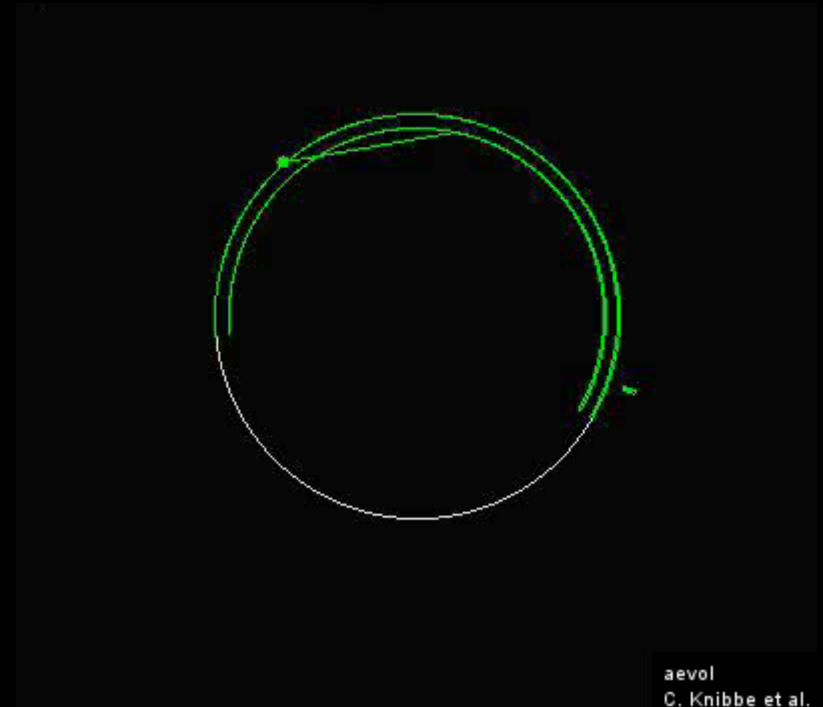
Generation = 0



High mutation rates : $2 \cdot 10^{-4}$ / pb

Genome length = 5000 bp

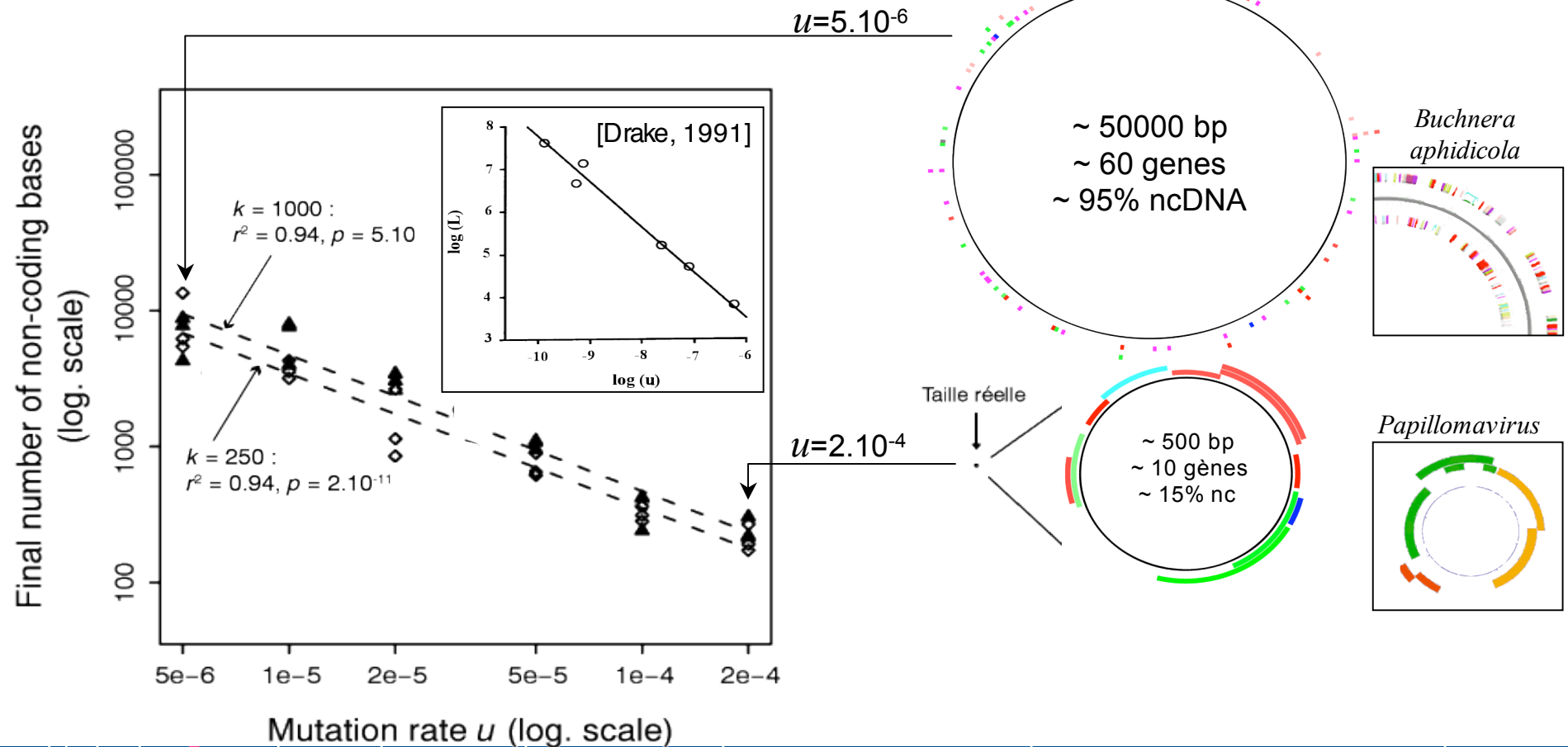
Generation = 0



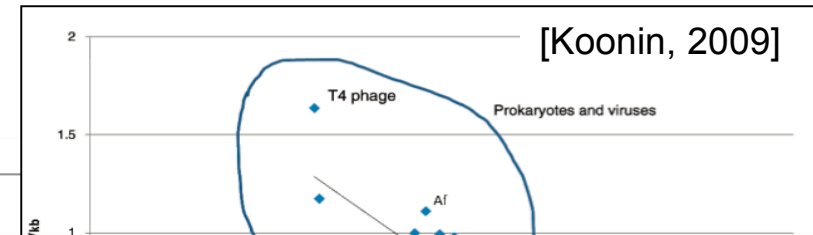
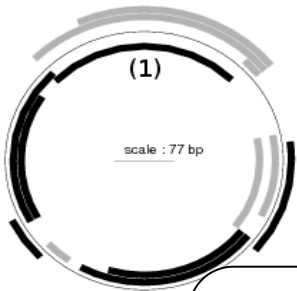
Low mutation rates : $5 \cdot 10^{-6}$ / pb

In silico experimental evolution

- Experimental study of the effect of mutation/rearrangement rates (u) and selection strength (k) on genome structure

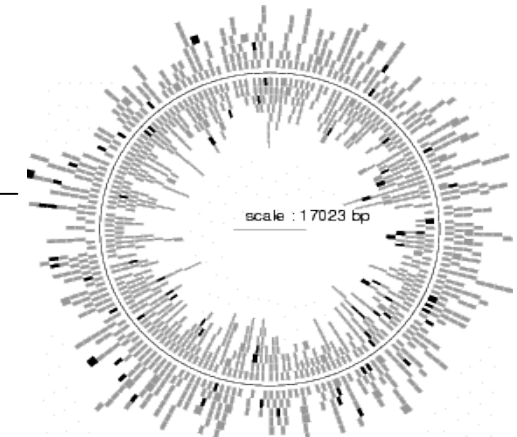
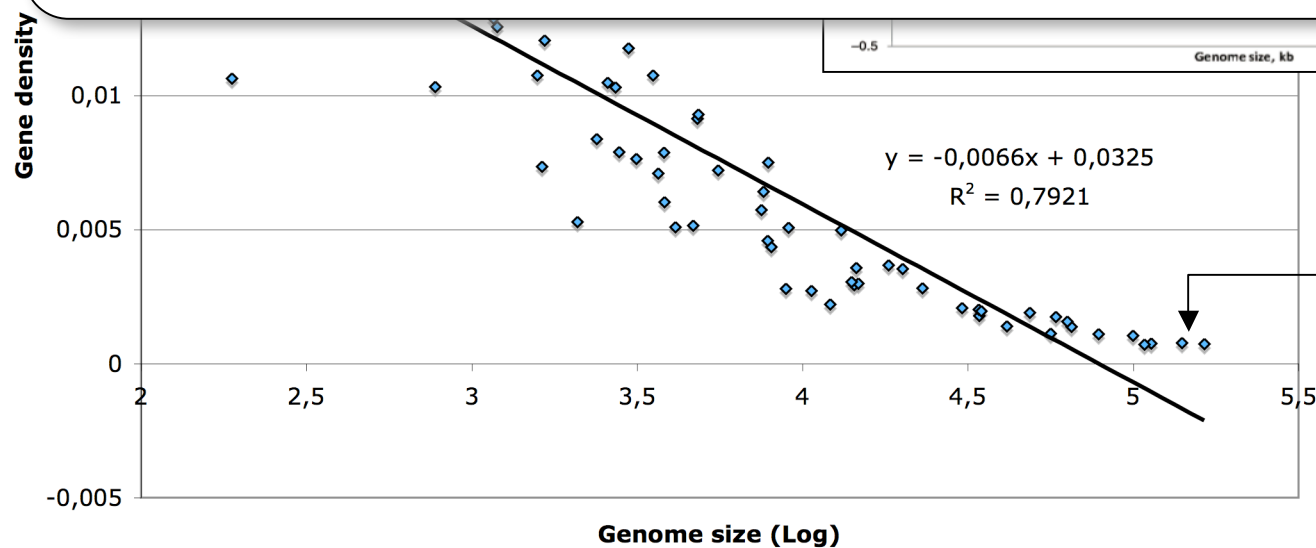


Yet another model explaining everything ;)



The model is able to reproduce known (but unexplained) data ...

But “*Prédire n’est pas expliquer*” (R. Thom) ...



[Parsons et al., Alife, 2010]

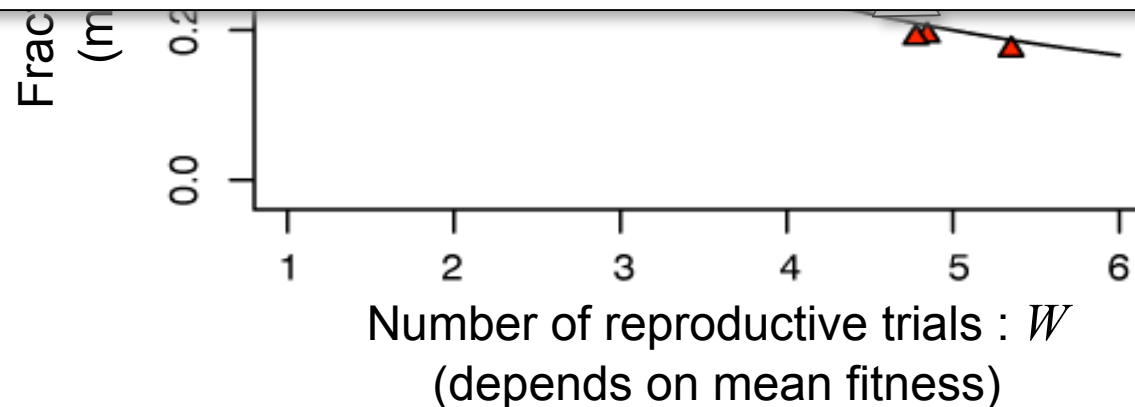
Measure of best individual reproduction neutrality (at $t = 20000$)

11



The regulation of the number of neutral offspring is the hallmark of an indirect selection process; the link between the mutation rate u and the size of the non-coding sequences show that the indirect selection depends (at least partly) on these sequences...

... But what is the link? Where does the burden come from?



Modeling the model ...

- Mathematical model of reproduction

- The math model is “true” for aevol *AND* for the “real world”...

- E*: Probability of neutral reproduction as a function of genome size (*L*),

If: (i) genomes undergo large duplications and deletions, (ii) the number and the average size of these events increase with genome size, Then: the mutational variability of a lineage depends on the amount of non-coding DNA (it is mutagenic for the genes it surrounds).

Thus the indirect selection for an appropriate level of variability actually selects for a specific amount of non-coding DNA

[Knibbe et al., Mol. Biol. Evol., 2007]

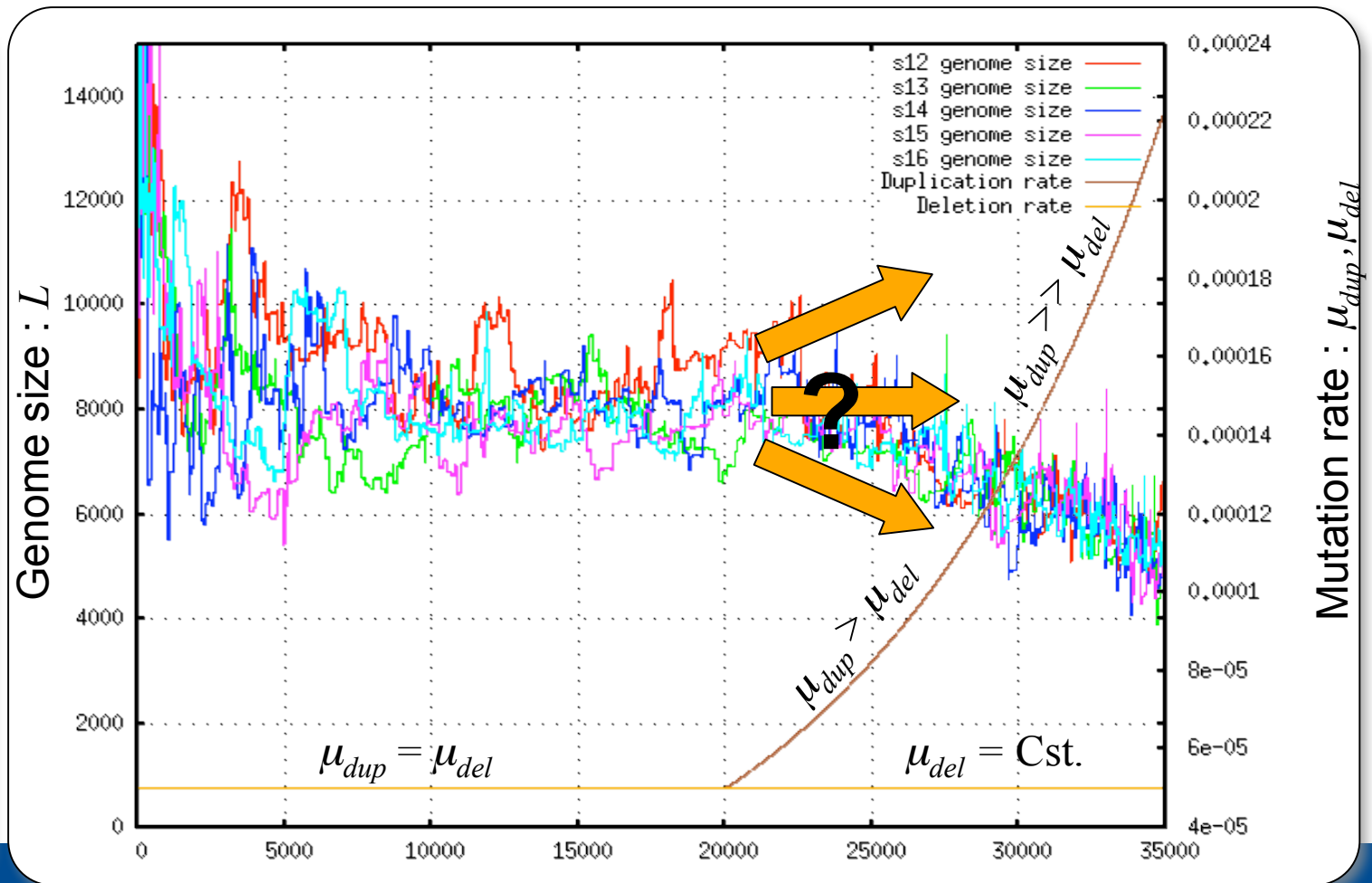
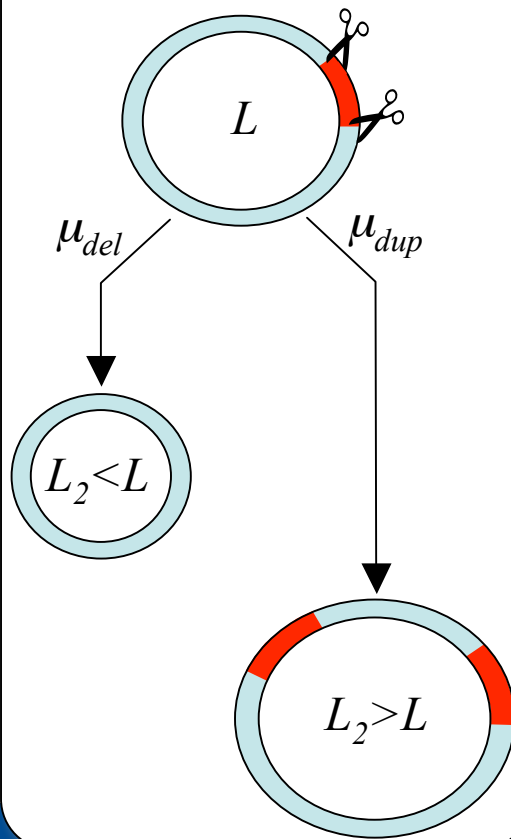
$$\left\{ \begin{array}{l} \tilde{\nu}_{\text{punct}} = \tilde{\nu}_{\text{ins}} = \tilde{\nu}_{\text{del}} = 1 - \frac{l}{L} \\ \tilde{\nu}_{\text{inv}} = \left(1 - \frac{l}{L}\right)^2 \\ \tilde{\nu}_{\text{transloc}} = \left(1 - \frac{l}{L}\right)^3 \end{array} \right\} \left\{ \begin{array}{l} \nu_{\text{gdel}} = \frac{1}{2L^2} \sum_{j=1} \lambda_j (\lambda_j + 1) \\ \tilde{\nu}_{\text{dup}} = \frac{1}{2L^2} \left(1 - \frac{l}{L}\right) \sum_{j=1}^{N_G} \lambda_j (\lambda_j + 1) \end{array} \right.$$



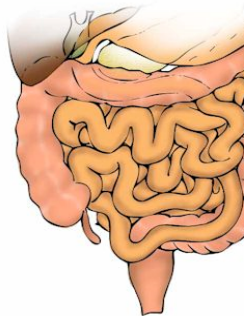
Surprising consequences

« It is simply a truism that the observed genome size is the result of a balance between the rate of DNA gain and loss » (Gregory, 2004)

DNA gain: duplications
DNA loss: deletions



What about gene networks?



Buchnera aphidicola str. APS (*Acyrtosiphon pisum*), complete...



Buchnera aphidicola

1 bp
genes
transcription factors

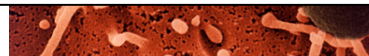
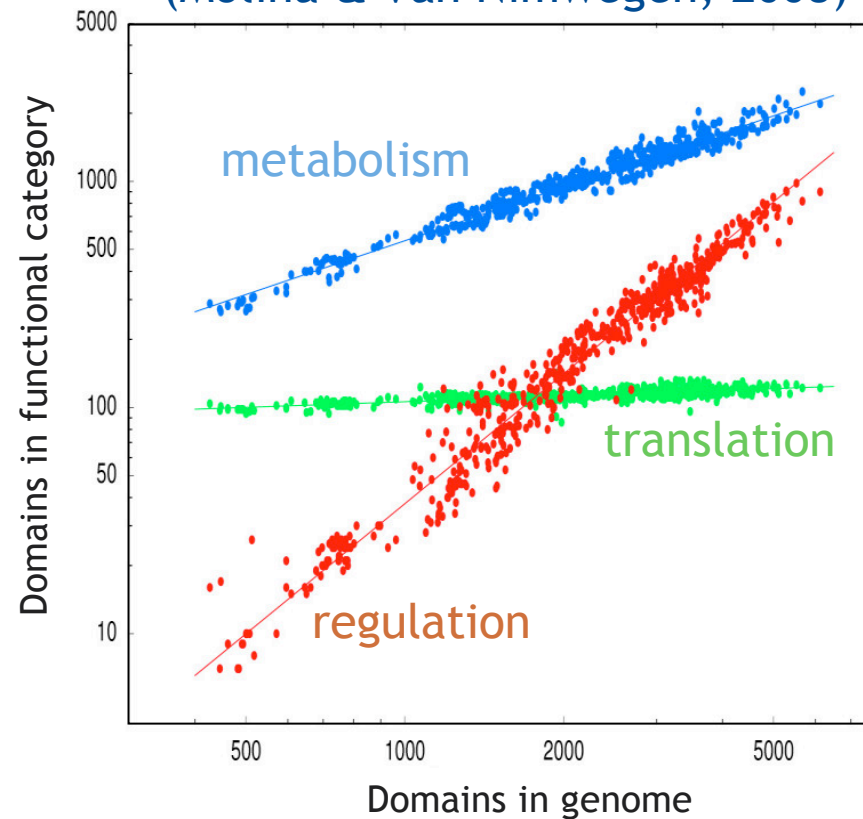
Escherichia coli

46 bp
genes
transcription factors

Escherichia coli

675 bp
genes
275 transcription factors

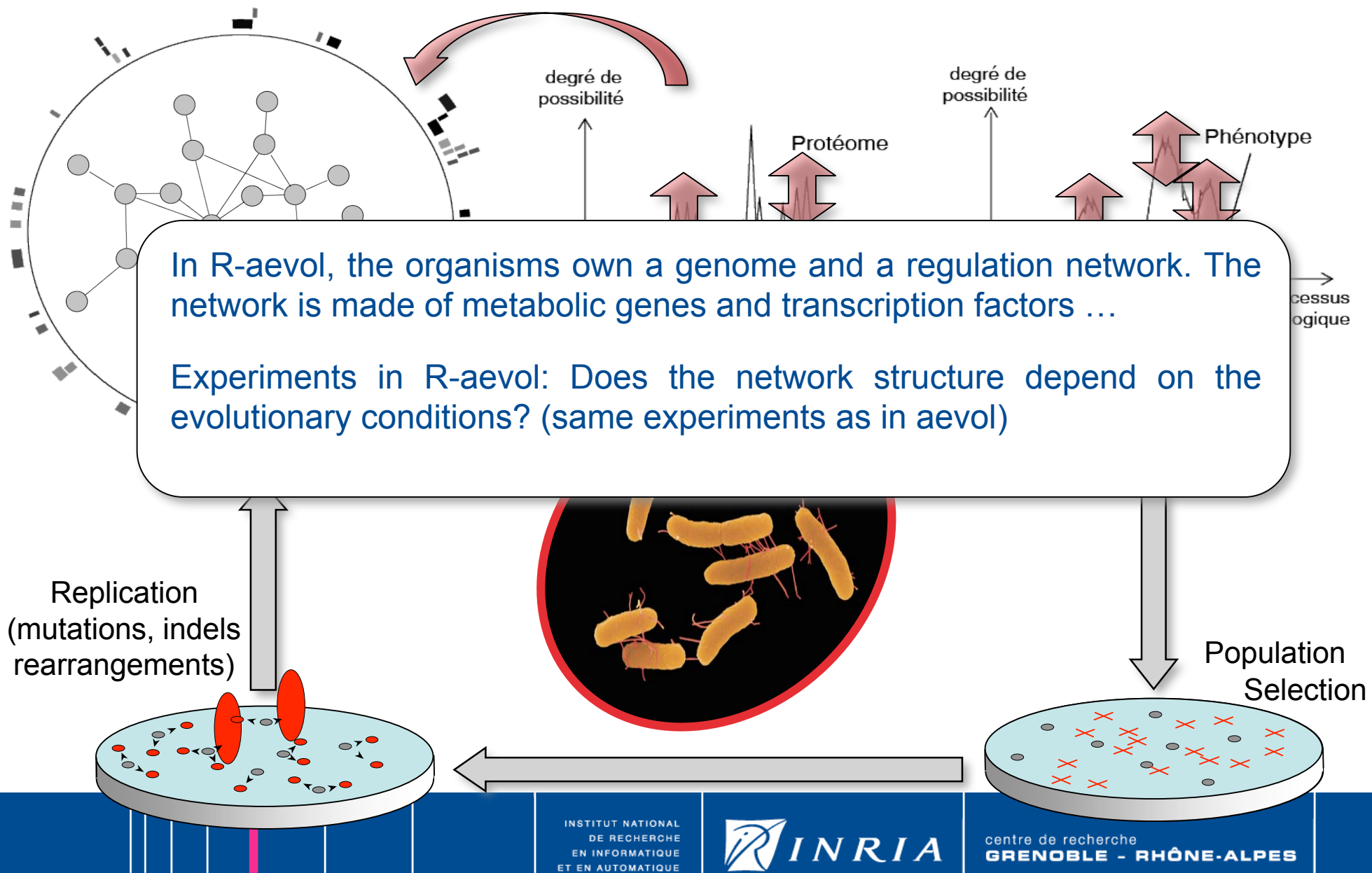
(Molina & Van Nimwegen, 2008)



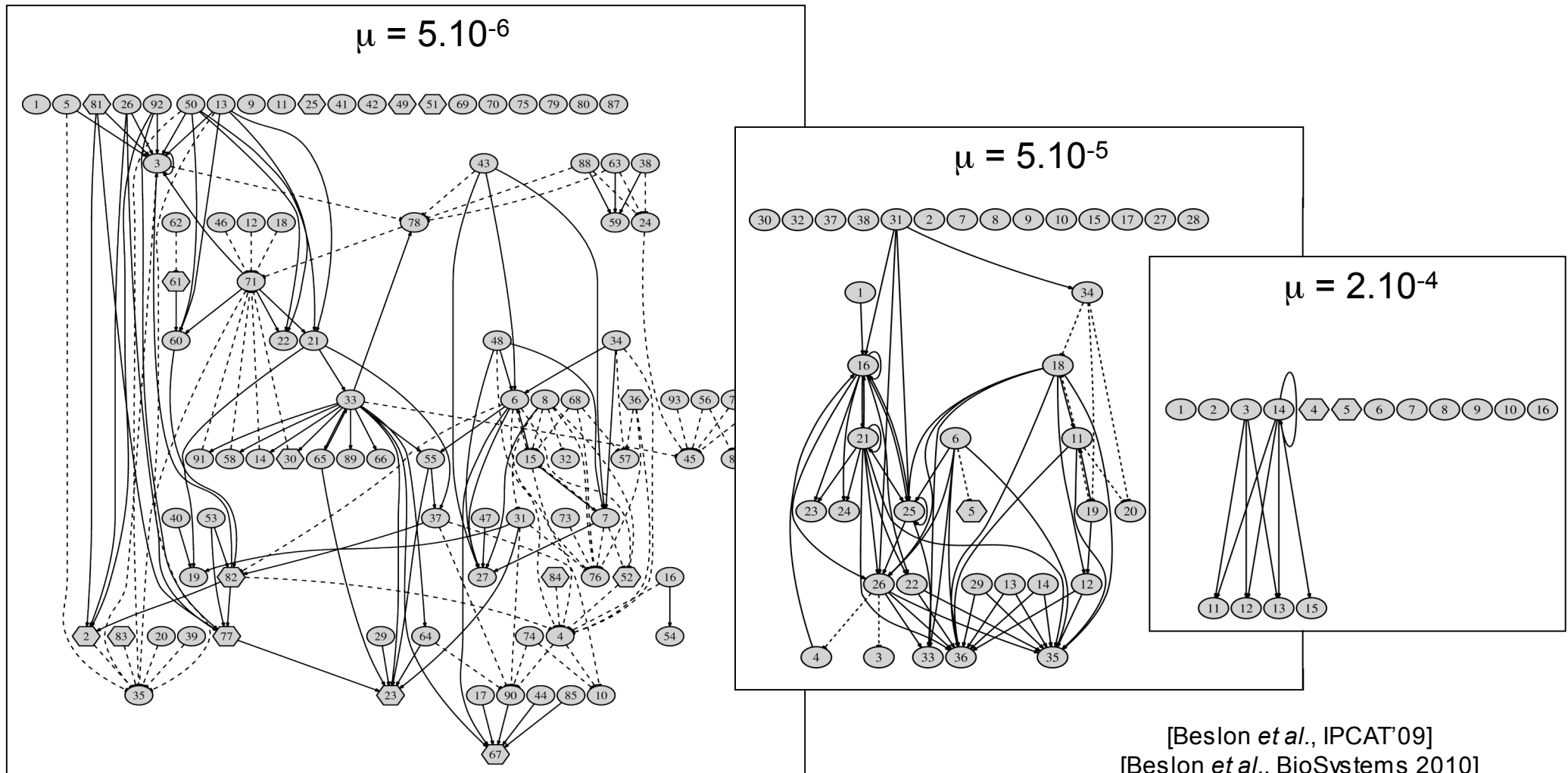
BA05: Friday April 15 09:42:20 2005

Length: 4,639,675 bp; Genes: 4,254

R-aevol (Y. Sanchez-Dehesa, 2009)

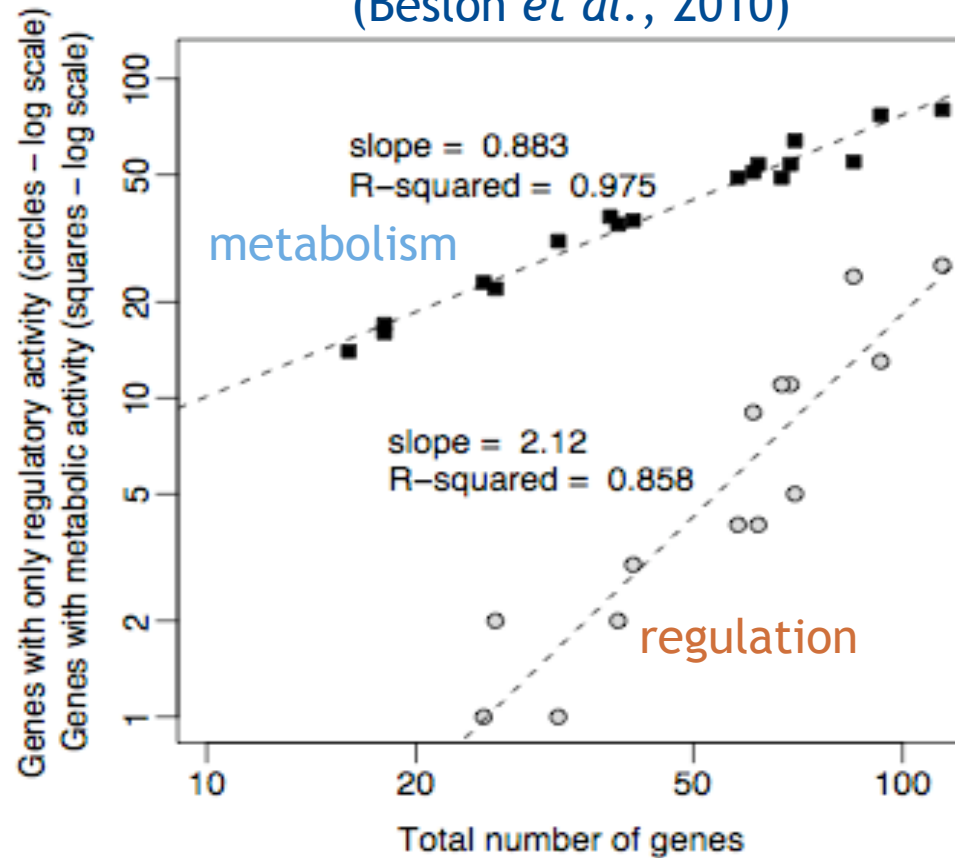


Impact of mutation rates on transcriptomic structures

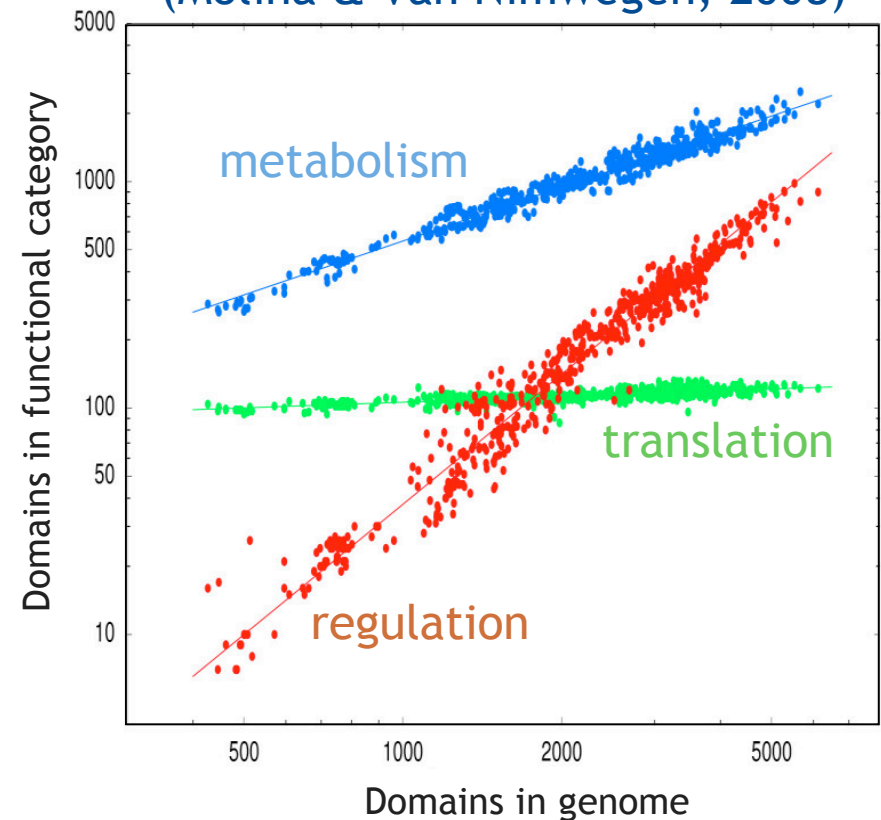


R-aevol: emergence of scaling laws

R-aevol
(Beslon *et al.*, 2010)



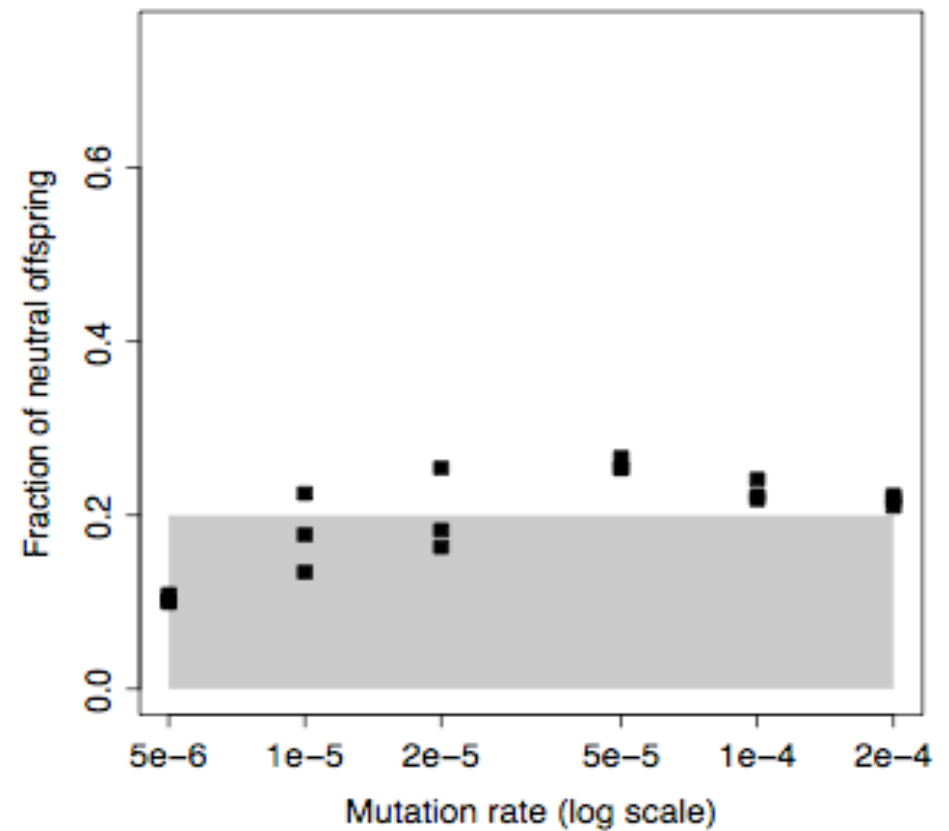
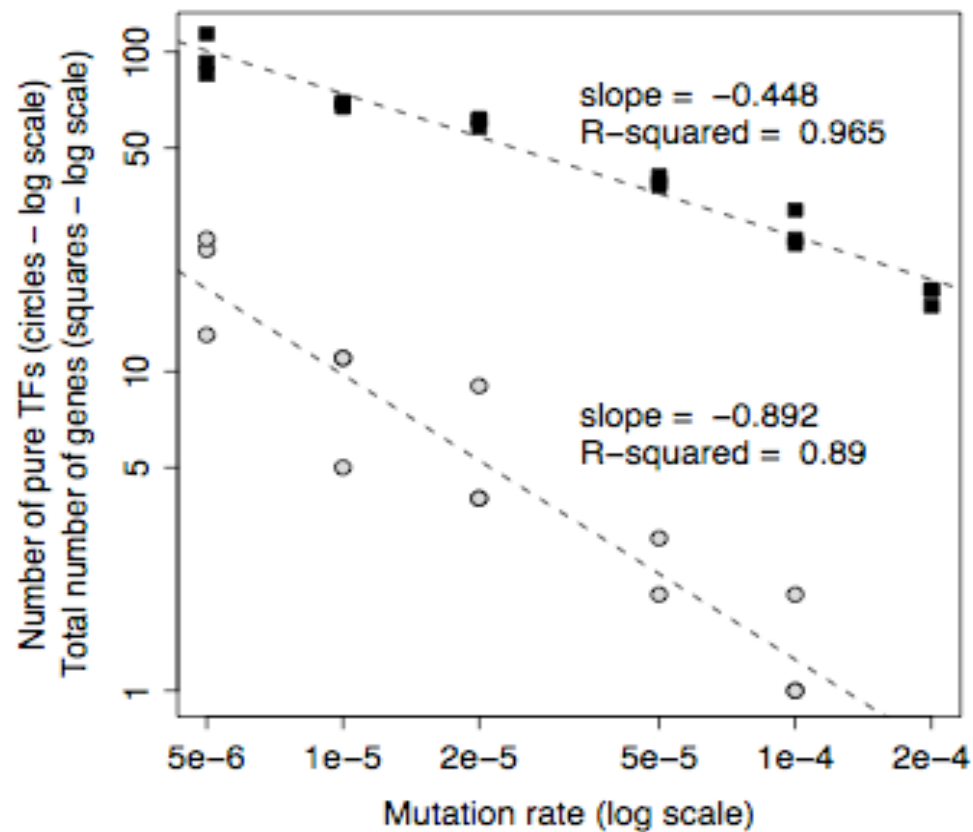
Biological data
(Molina & Van Nimwegen, 2008)



Side effect of the selection for robustness?

R-aevol

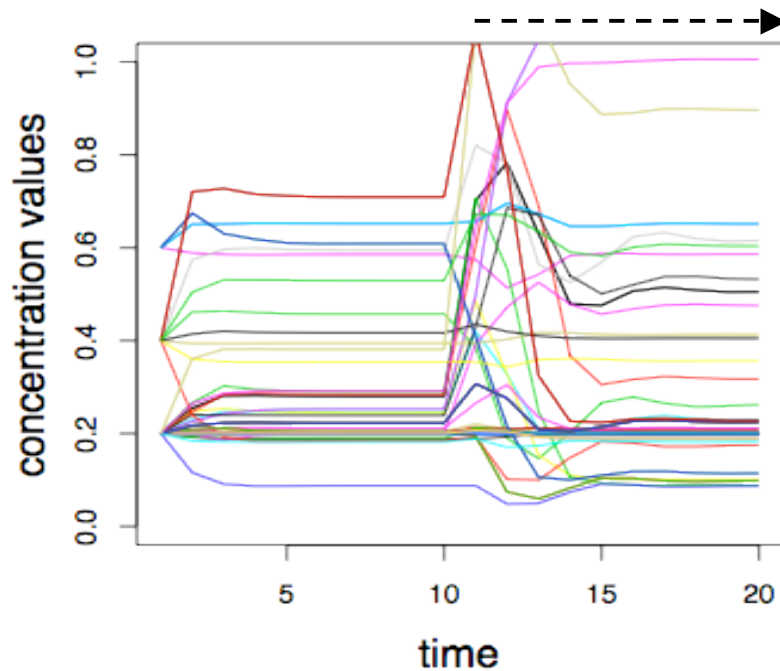
$F_v \sim \text{constant}$



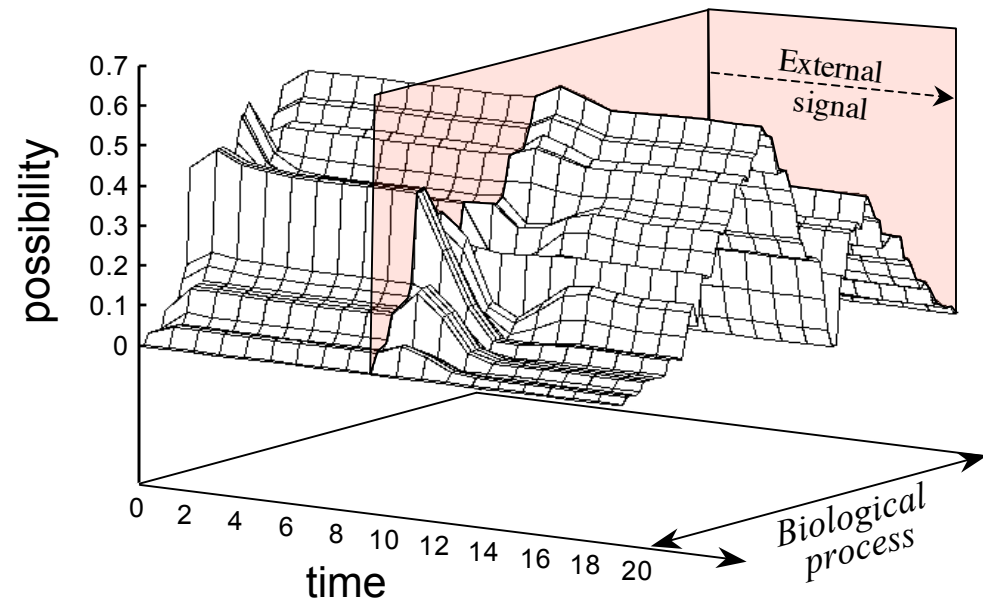
What if the organisms have “something to do”

- Organisms live for 20 time steps ; at $t = 10$ a signal is sent to the “cells” that must react by changing their phenotypes...

Protein concentration over time

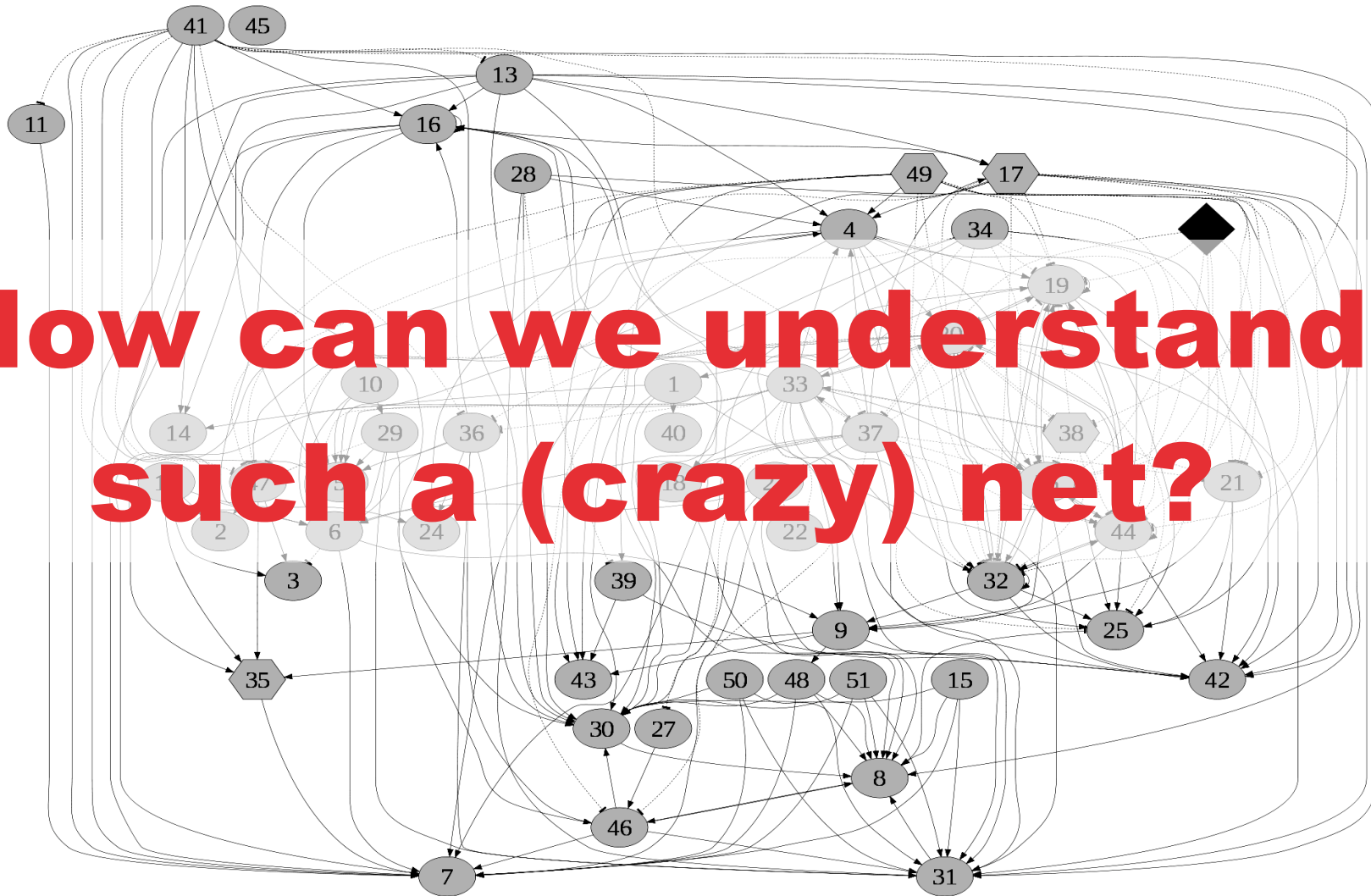


Phenotype over time

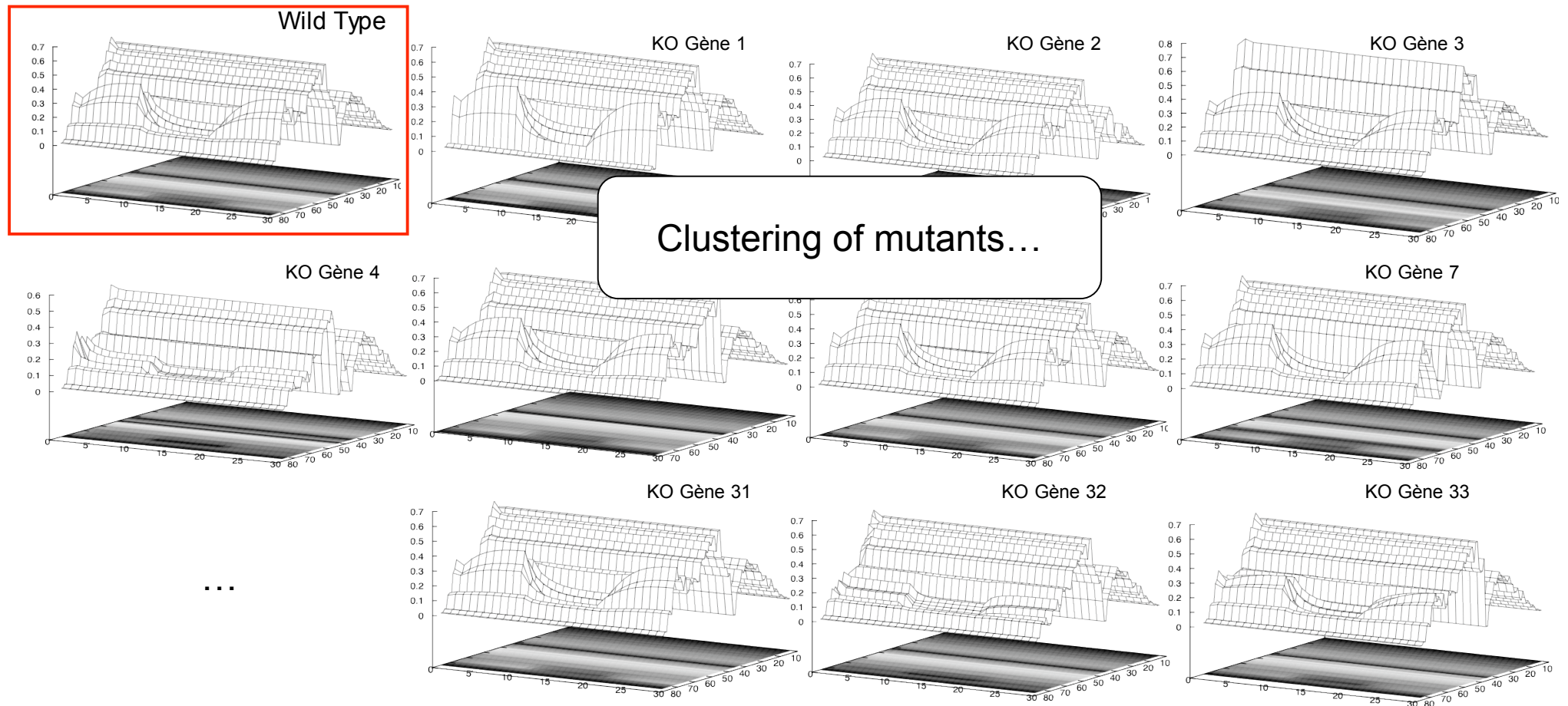


Evolved network after 15000 generations

**How can we understand
such a (crazy) net?**



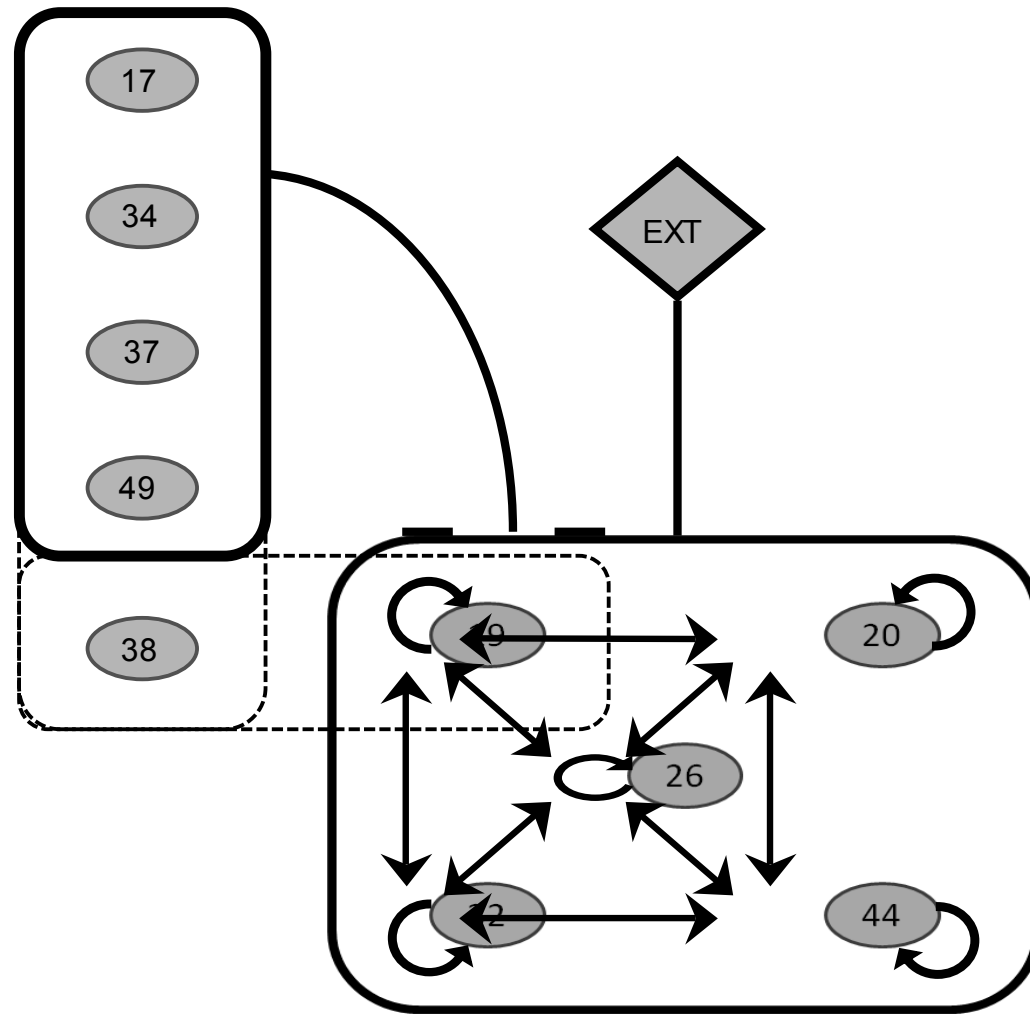
Systematic Knock-Out experiments



[Beslon et al., IDA], 2010]



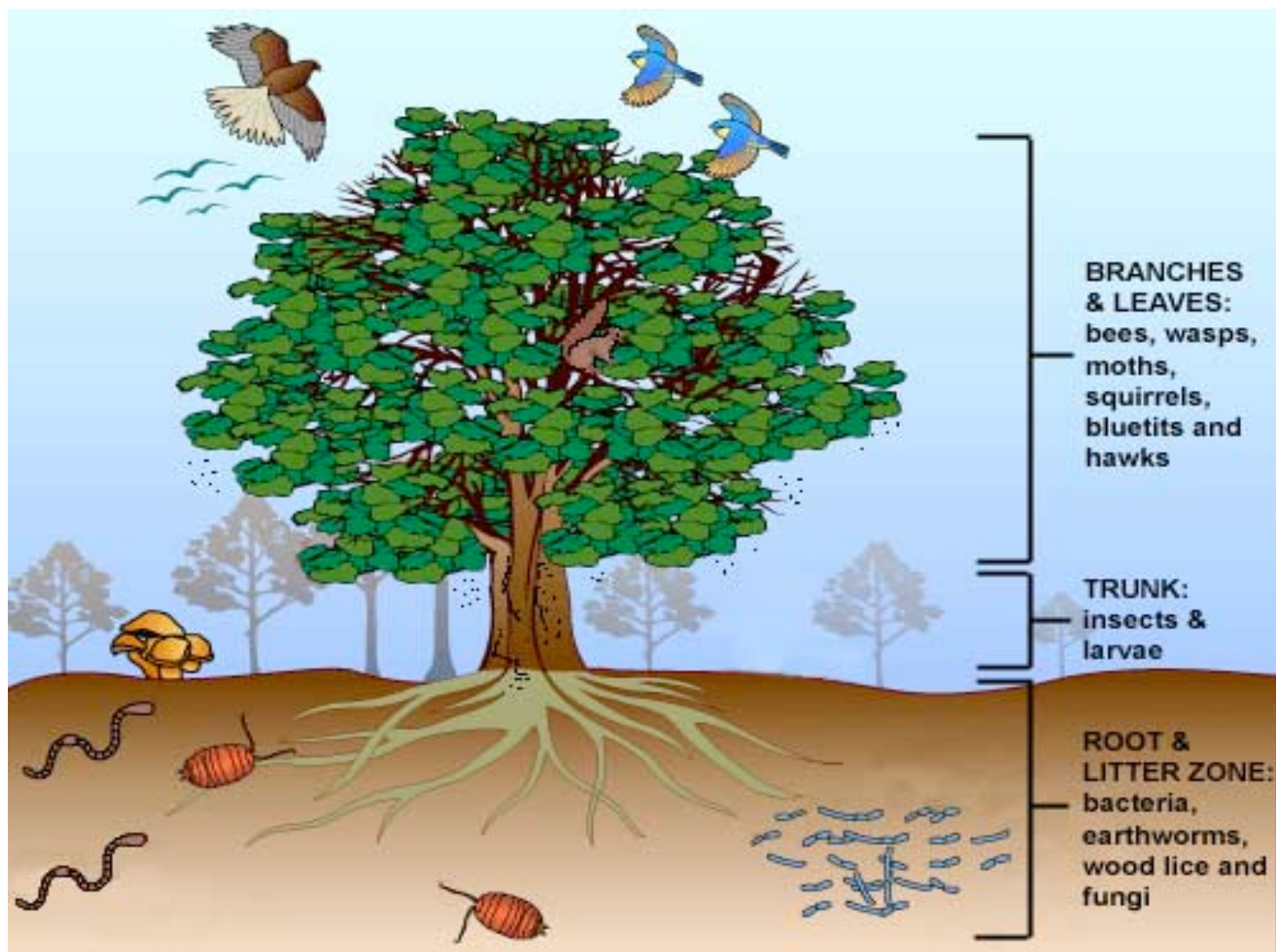
Network sketch with two modules



Origin of transcriptomics complexity?

- Where does the network complexity come from?
 - [In less stable, more changing environments, transcription factors are over-represented] ... *This suggests that in ever-changing, highly competitive environments, there is a strong selective pressure towards regulated and coordinated gene expression, compared with very stable environments. (Cases et al., 2003)*
- According to this view, the origin of (transcriptomic) complexity is another complexity (environmental)!
 - But in our experiments, the complex network emerges in a simple environment (one stable state) as well as in two-states environments
- Thus complexity emerges “for free” (at least in the model)
 - Environmental complexity is NOT a necessary condition
 - A new analysis paradigm for genetic networks understanding?





References

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